Elongation and Fluctuations of Semiflexible Polymers in a Nematic Solvent


1Department of Physics and Astronomy, University of Pennsylvania, Philadelphia, Pennsylvania 19104, USA
2Department of Chemical and Biomolecular Engineering, University of Pennsylvania, Philadelphia, Pennsylvania 19104, USA
3Institute for Medicine and Engineering, University of Pennsylvania, Philadelphia, Pennsylvania 19104, USA

(Received 13 October 2003; published 24 March 2004)

DOI: 10.1103/PhysRevLett.92.125503 PACS numbers: 61.30.-v, 64.70.Md, 82.35.Pq

We directly visualize single polymers with persistence lengths \( \ell_p \), ranging from 0.05 to 16 \( \mu m \), dissolved in the nematic phase of rodlike \( fd \) virus. Polymers with a sufficiently large persistence length undergo a coil-rod transition at the isotropic-nematic transition of the background solvent. We quantitatively analyze the transverse fluctuations of the semiflexible polymers and show that at long wavelengths they are driven by the fluctuating nematic background. We extract the Odijk deflection length and the elastic constant of the background nematic phase from the data.

Polymer coils in solution exhibit a variety of conformational and dynamical behaviors depending on many factors, including polymer concentration, polymer stiffness, solvent quality, solvent flow, and mechanical stress. Exciting recent experiments in this field have focused on disentanglement of single biopolymers in isotropic solutions as a result of applied forces and solvent flow [1], and on transport of single biopolymers through networks of barriers [2]. In this Letter, we explore conformations of polymer coils in anisotropic solutions. In particular, we present the first direct experimental observations of isolated semiflexible polymers dissolved in a background nematic phase composed of aligned rodlike macromolecules. We show by direct visualization that semiflexible biopolymers dissolved in the nematic phase assume an elongated rodlike configuration aligned with the background nematic director. A coil-rod transition of the biopolymer can thus be induced by causing the solvent to undergo an isotropic-to-nematic (I-N) transition. We quantitatively explore the fluctuations of these semiflexible polymers and find they cannot be described by a theory which treats the nematic background as a fixed external field [3].

Mixtures of semiflexible polymers in lyotropic nematic suspensions exemplify an emerging class of complex fluids — hypercomplex fluids, such as nematic elastomers [4] and nematic emulsions [5], wherein two or more distinct components are combined to create systems that exhibit novel physical properties and functions. Understanding of the polymer-nematic system may lead to new ideas about how to achieve high alignment of biopolymers, complementary to existing methods of DNA alignment [6]. Furthermore, since many biopolymers such as the actin filaments within the sarcomere and neurofilaments within the axon reside in an anisotropic, nematic-like environment [7], our investigation may shed light on the actin filaments within the sarcomere and neurofilaments [6].

Our experiments employ an aqueous solution of rodlike \( fd \) viruses as a background nematic liquid crystal. This system has been studied extensively [12–14], and its phase behavior is well described by the Onsager theory for rods with hard core repulsion [15]. Another advantage of this system is its compatibility with most biopolymers.

We use four different semiflexible polymers, whose physical parameters are listed in Table I. To directly visualize the polymers dissolved in the nematic background, we fluorescently labeled each polymer: DNA was labeled with YOYO-1 (Molecular Probes, Eugene, OR), neurofilaments with succinimidyl rhodamine B [20], F-actin filaments with rhodamine-phallolidin (Sigma, St. Louis, MO), and wormlike micelles with PKH26 dye (Sigma, St. Louis, MO) which preferentially partitions into the hydrophobic core of the micelle. Since DNA, neurofilaments, and actin are all negatively charged, we expect

<table>
<thead>
<tr>
<th>Polymer</th>
<th>( L (\mu m) )</th>
<th>( \ell_p (\mu m) )</th>
<th>( a (nm) )</th>
<th>Ref.</th>
</tr>
</thead>
<tbody>
<tr>
<td>A-DNA</td>
<td>16</td>
<td>0.05</td>
<td>2</td>
<td>[16]</td>
</tr>
<tr>
<td>Neurofilament</td>
<td>2–10</td>
<td>0.2</td>
<td>10</td>
<td>[17]</td>
</tr>
<tr>
<td>Wormlike Micelles</td>
<td>5–50</td>
<td>0.5</td>
<td>14</td>
<td>[18]</td>
</tr>
<tr>
<td>F-actin</td>
<td>2–20</td>
<td>16</td>
<td>7</td>
<td>[19]</td>
</tr>
<tr>
<td>( fd ) virus</td>
<td>0.9</td>
<td>2.2</td>
<td>7</td>
<td>[13]</td>
</tr>
</tbody>
</table>

TABLE I. The contour length \( L \), the persistence length \( \ell_p \), and the diameter \( a \) of biopolymers in our experiments.
them to be stable in a suspension of negatively charged *fd* viruses. Wormlike micelles are sterically stabilized with a neutral poly(ethylene oxide) brush layer, which does not interact with the *fd* virus or other proteins [13,18].

Bacteriophage *fd* was grown and dialyzed against a phosphate buffer (150 mM KCl, 20 mM phosphate, 2 mM MgCl₂, pH = 7.0) [13]. Samples were prepared by mixing a small amount of polymer with a *fd* solution at different concentrations and were placed between a coverslip and a glass slide. A chamber with a thickness of ~50 μm was made using a stretched parafilm as a spacer. Samples sealed with optical glue (Norland Products, Cranbury, NJ) were allowed to equilibrate until no drift was visually detectable. To reduce photobleaching, we added an antioxygen solution (2 mg/ml glucose, 360 U/ml catalase, 0.25 vol % mercaptoethanol, 8 U/ml glucose oxidase). All samples were imaged with a fluorescence microscope (Leica IRBE) equipped with a 100× oil-immersion objective and a 100 W mercury lamp. Images were taken with a cooled CCD camera (CoolSnap HQ, Roper Scientific), which was focused at least 5 μm away from the surface to minimize possible wall effects.

Figure 1 displays a series of pictures that summarize our qualitative observations. In the nematic phase of *fd*, F-actin [Fig. 1(a)], wormlike micelles [Fig. 1(b)], and neurofilaments [Fig. 1(c)] are highly elongated, having a rodlike shape. By contrast, the same filaments dissolved in an isotropic phase crumple into more compact random coils. Just above the I-N transition, actin filaments and wormlike micelles form hairpin defects [9]. These hairpins exhibit interesting dynamics as shown in Fig. 1(e) and will be explored by us elsewhere. DNA dissolved in *fd* nematic behaves qualitatively differently [Fig. 1(d)]; it forms a slightly anisotropic droplet. Each droplet contains many DNA molecules and, with time, these droplets coalesce into larger droplets. Thus, even at a very low concentration, DNA separates from the *fd* nematic. Taken together, these observations suggest that the persistence length of the polymer is important in determining its solubility in the nematic liquid crystals: DNA has a small *lp* and is insoluble, unlike the other stiffer polymers in our experiments. This insolubility may be related to the entropy-driven phase separation of a system of bidisperse rigid rods if their lengths and/or diameters are sufficiently dissimilar [21]. This theory, however, has not been extended to the case of semiflexible polymers.

The large contour lengths of actin filaments and wormlike micelles make them suitable for further quantitative analysis. We focus on the fluctuations of filaments in a background nematic that is free of both defects and distortions. A series of 50 to 100 images were taken with a few seconds between each image to ensure that statistically independent configurations were sampled. For each *fd* concentration, ten filaments were analyzed. The conformation of each polymer was reconstructed by manually marking the end points. [Note that we parametrize...]

**FIG. 1.** Images of fluorescently labeled biopolymers in the isotropic (left panels) and nematic (right panels) phases of the *fd* virus. (a)–(d) are, respectively, the images of actin, wormlike micelles, neurofilaments, and DNA. The polymers in an isotropic solution are confined by a thin chamber thus making the samples quasi two dimensional. (e) A sequence of images illustrating an actin filament escaping from a hairpin defect. The scale bar is 5 μm. (f) A schematic of a biopolymer in the background nematic field; the conformation of the polymer is parametrized by \( \mathbf{R}(z) = (r_x(z), r_y(z), z) \). The nematic director points along the \( z \) axis.

**FIG. 2.** The order parameter of actin filaments (\( S_{\text{actin}} \)) vs the order parameter of the background *fd* nematic (\( S_{\text{fd}} \)). Dashed line is a guide to the eye. The contour length of actin filaments is 15 μm or higher. The values of \( S_{\text{fd}} \) at different *fd* concentrations have been measured in Ref. [14]. Inset: The orientation distribution function (ODF) of actin filaments. The ODF is well approximated by a Gaussian distribution for a wide range of concentrations.
the transverse deviations of the polymer from the \( z \) axis by the 2-component vector \( \mathbf{r}(z) \), as shown in Fig. 1(f).] An intensity profile along the \( x \) direction for each value of \( z \) was extracted. By fitting this intensity to a Gaussian, we obtained subpixel accuracy for \( r_s(z) \). We first extracted the orientational distribution function (ODF) from our data. Since our images are two-dimensional projections of the polymer fluctuating in three dimensions, the \( x \) component of the tangent vector is measured by \( t_x(z) = \partial r_s(z)/\partial z \simeq \theta_z(z) \). The ODF is obtained by creating a histogram of \( \theta_z \) at different positions along the contour length for a time sequence of 50–100 images. A typical ODF is plotted in Fig. 2; it is well approximated by a Gaussian distribution.

Next, we compute the order parameter of the polymer defined by \( S = \int_0^L dz \langle 3\mathbf{t}(z) \cdot \mathbf{t}(z) \rangle - 1/(2L) = 1-3\langle t_z^2(0) \rangle \), where in the last equality, we have used \( \langle t_z^2(z) \rangle = \langle t_z^2(0) \rangle \). In Fig. 2, we plot \( S \) for actin as a function of the background nematic order parameter. It is interesting to observe that \( S_{\text{actin}} \) is significantly higher than \( S_{\text{fd}} \). In order to check that the difference in the alignment between actin and \( \text{fd} \) molecules is due to their different contour lengths, we measured \( S_{\text{actin}} \) for different contour lengths of actin filaments, as shown in Fig. 3. As the contour length of actin decreases, \( S_{\text{actin}} \) approaches \( S_{\text{fd}} \), as expected intuitively. These observations are qualitatively consistent with the Onsager theory for a bidisperse mixture of rodlike particles with different lengths considered in Ref. [22]. This theory predicts the order parameter of long rods will be higher than the order parameter of the background nematic of shorter rods.

\[
\langle t_x(z') t_x(z' + z) \rangle = \frac{\lambda}{4 \ell_p} e^{-z/\lambda} + \frac{1}{8 \pi^2 K \lambda} \int_0^\infty dx \frac{\cos(xz/\lambda) \log(1 + D^2/x^2)}{(1 + x^2)(1 + 2 + 4\pi K \log(1 + D^2/x^2))},
\]

Finally, we measured the tangent-tangent correlation function (TTCF) \( \langle t_x(z') t_x(z' + z) \rangle \) for wormlike micelles dissolved in the \( \text{fd} \) virus with concentration 40 mg/ml and larger (Fig. 4). At low \( \text{fd} \) concentrations, the fluctuations of worms are large as evidenced by visual observation of spontaneous formation and dissolution of hairpin defects. In this regime, the measured TTCF does not decay uniformly. We thus focus our analysis on the regime where the background order parameter is very high, and the amplitude of the polymer fluctuations is small. This makes our data suitable for comparison with the theoretical model outlined below.

The fluctuations of a semiflexible polymer in a nematic phase may be described by the free energy [10,24]:

\[
F = \frac{k_B T}{2} \int_0^L dz \left\{ \ell_p \left[ \frac{\partial \mathbf{t}}{\partial z} \right]^2 + \Gamma [\mathbf{t} \cdot (\nabla \delta \mathbf{n})]^2 \right\} + \frac{1}{2} \int d^3 x K (\nabla \delta \mathbf{n})^2,
\]

where \( k_p \) is the Boltzmann constant, \( T \) is the temperature, \( \ell_p \) is the persistence length of the semiflexible polymer, \( \Gamma \) is the strength of the coupling of the polymer to the background nematic field, \( \delta \mathbf{n} \) is the local direction of the fluctuating nematic field, and \( K \) is the nematic elastic constant. Note that \( \mathbf{t} \) and \( \delta \mathbf{n} \) are two-dimensional vectors in the plane perpendicular to the average director. It is straightforward to compute \( \langle t_x(z') t_x(z' + z) \rangle \) from Eq. (1):
where $D = 2 \pi \lambda / a$ is related to the molecular cutoff which we assume to be the diameter of the polymer ($a \sim 10 \text{ nm}$). The first term in Eq. (2) describes the fluctuations of a semiflexible polymer in a static external field, with a decaying length set by $\lambda = \sqrt{\ell_p / \Gamma}$. The second term describes the fluctuations of the polymers driven by the tight coupling to the fluctuations of the background nematic field. This term generalizes that of Ref. [10], since it includes the back reaction of the stiff polymer on the nematic fluctuations. Because it decays approximately as a power law, we expect at large length scales the fluctuations of the polymer are always dominated by the nematic fluctuations.

Figure 4 shows our measured TTCF along with the fitted curve of Eq. (2). Overall, good agreement is obtained at distances above $0.5 \mu m$ [23]. At these distances, most of the fluctuations of the worms are driven by the tight coupling to the background nematic field, coming from the second term in Eq. (2). The best-fit value of $\ell_p$ is found to be $1.5 \mu m$, somewhat higher than those obtained in previous measurements [18]. From the fits to the data, we extract values of the Odijk deflection length $\lambda$, $K$, and $\Gamma$ as listed in Table II. We observe that with increasing $fd$ concentration, $\lambda$ decreases, while $K$ and $\Gamma$ increase, as one would intuitively expect. Finally, we note that the values for $K$ are in agreement with previous measurements of twist elastic constant $K_{22} = 3 \times 10^{-8} \text{ dyn for } fd$ samples prepared under similar conditions [25].

In conclusion, we have shown that semiflexible polymers with large enough persistence lengths assume a rodlike conformation when dissolved in a nematic solvent. Using image analysis, a full nematic orientational distribution function was measured. In addition, we have shown that fluctuations of the polymer are driven primarily by the fluctuations of the background nematic field. Direct visualization of individual polymer yields valuable new information about the behavior of polymer chains in anisotropic solvents.

This work was supported by the NSF through Grant No. DMR-0203378 (A. G. Y.), DMR01-29804 (R. D. K.), MRSEC Grant No. DMR-0079909, NASA NAG8-2172 (A. G. Y.), NIH R01 HL67286 (P. A. J.), and the Donors of the Petroleum Research Fund, administered by the American Chemical Society (R. D. K.).

*Current address: Rowland Institute at Harvard, Cambridge, MA 02142, USA.

[23] At distances smaller than $0.5 \mu m$, we observe a significant deviation of our data from the theoretical curve, which likely arises from the limited spatiotemporal resolution of our microscope. Over the image acquisition time of 250 ms, the fast fluctuations of the polymers at short wavelengths are effectively washed out, leading to a lower value in the TTCF. Another possible source of discrepancy is that the spatial resolution of the microscope is smaller than the Odijk deflection length.